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Development of the first and practical method for enantioselective synthesis of ¹⁰B-enriched *p*-borono-L-phenylalanine

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1. Introduction

Boron neutron capture therapy (BNCT) based on the selective accumulation of ¹⁰B compounds in tumor cells and the subsequent irradiation with thermal neutron is highly noted as one of useful techniques for treatment of cancer.^{1–3} So far *p*-(¹⁰B)boronophenylalanine (¹⁰Bpa) (1) in which boron atom is enriched with ¹⁰B isotope had been developed as an excellent ¹⁰B carrier for BNCT (Fig. 1). Since the incorporating amount of L-¹⁰Bpa into cancer cells is higher than that of D-¹⁰Bpa or DL-¹⁰Bpa is now being used clinically for the treatment of patients with malignant brain tumor and/or melanoma owing to the selective accumulation and low toxicity.⁵ At present, commercially available L-¹⁰Bpa is prepared by the enzymatic resolution of the racemic precursor.⁶

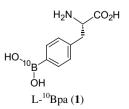


Figure 1. The structure of L-¹⁰Bpa (1).

ABSTRACT

At present p-(¹⁰B)borono-L-phenylalanine (L-¹⁰Bpa) is used clinically as an excellent ¹⁰B carrier for BNCT; however, its enantioselective synthesis has not been reported yet. The present Letter describes the first and practical method for enantioselective synthesis of ¹⁰B-enriched L-Bpa using Z-L-Ser–OMe as the chiral synthon in good total yield and high optical purity.

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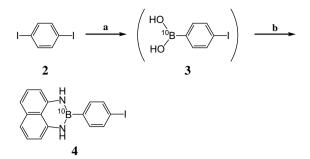
2. Results and discussion

To date, three synthetic approaches of L-Bpa with naturally occurring boron atom⁷ utilizing the enantioselective method have been known. For example, Kirihata and co-workers reported the enantioselective synthesis of L-Bpa based on the coupling reaction of 4-boronobenzylbromide with the chiral derivative prepared from L-valine.⁸ However, this method needs further enzymatic resolution to obtain optically pure L-Bpa, since the enantioselectivity of the coupling reaction is slightly low. Furthermore, Yamamoto and co-workers reported the method based on palladium catalyzed coupling reaction of pinacolborane with 4-iodo-L-phenylalanine derivatives.⁹ This method is also not suitable for the synthesis of L-¹⁰Bpa, because ¹⁰B-enriched pinacolborane is not commercially available and its synthetic method is not reported as well. In another case, Malan and Morin reported an interesting method utilizing the Negishi coupling reaction of iodophenylborate and L-iodoalanine derivatives.¹⁰ However, this method is also unsuitable for the synthesis of L-¹⁰Bpa, because both ¹⁰B-enriched BI₃ and 1,3-diphenylpropane-1,3-diol used as the protecting group of boron are not commercially available. Consequently, all these methods are not applicable to the synthesis of ¹⁰B-enriched L-Bpa.

Therefore, we examined a development of efficient method for enantioselective synthesis of L-¹⁰Bpa utilizing the Negishi reaction based on the coupling of 4-(¹⁰B)borono-iodobenzene derivative with 3-iodo-L-alanine [L-Ala(3I)] derivative. The most important advantage of the present method is that ¹⁰B(OMe)₃ as ¹⁰B source to prepare 4-(¹⁰B)boronoiodobenzene is commercially available. Further L-Ala(3I) can be readily prepared from L-serine. Thus, the synthesis of L-¹⁰Bpa was carried out as shown in Schemes 1 and 2.

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Scheme 1. Reagents and conditions: (a) (i) *n*-BuLi (1.00 equiv), THF, -78 °C, 30 min; (ii) $^{10}B(OMe)_{3}$, -78 °C, 5 h; (b) 1,8-diaminonaphtalene, THF, rt, 12 h, 57.7% (2 steps from **2**).

First, 1,4-diiodobenzene (**2**) was allowed to react with ${}^{10}B(OMe)_3$ to give 4-(${}^{10}B$)boronoiodobenzene (**3**). The borono group of **3** was then protected with 1,8-diaminonaphtalene that was developed as a masking agent in the Suzuki–Miyaura cross coupling reaction by Suginome and co-workers¹¹ to give 4-[2,3-dihydro-1*h*-naphtho[1,8-*de*]-1,3-diaza-2-(${}^{10}B$)borinyl]iodobenzene (**4**) (Scheme 1).

Next, Z-L-Ala(3I)-OMe (6)¹² was prepared from Z-L-Ser-OMe (5)¹³ (Scheme 2), and the compound 6 was coupled with the aryliodide **4** via Zn adduct **6**′ by employing the Negishi coupling reaction to give N^{α} -benzyloxycarbonyl-*p*-[2,3-dihydro-1*H*-naphtho[1,8-*de*]-1,3-diaza-2-(¹⁰B)borin-yl]-L-phenylalanine methyl ester (7). To find the optimal conditions for the Negishi coupling, we examined the relationship between the yields and the catalysts on various reaction conditions as shown in Table 1. The reaction scarcely proceeded in the presence of Pd(PPh₃)₂Cl₂ (entries 1 and 2) as a catalyst. When the reaction was carried out at 70 °C with Pd(PPh₃)₄ catalyst, L^{-10} Bpa derivative **7** was produced in a 42.7% yield (entry 4), although the reaction did not proceed at room temperature (entry 3). As a result, the reaction proceeded favorably at rt by the use of Pd₂(dba)₃ and P(o-tolyl)₃ as catalysts,¹⁴ although the yield was slightly decreased when the reaction was carried out at 70 °C (entries 5 and 6).

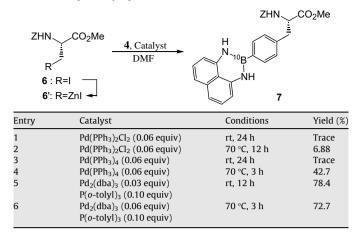
The diaminonaphthyl group of compound **7** was cleaved with 2 M H_2SO_4 in advance to give Z-L-¹⁰Bpa–OMe (**8**). The benzyloxycarbonyl and methyl groups were simultaneously removed by acid hydrolysis with 3 M HCl to give L-¹⁰Bpa (**1**) (Scheme 2). HPLC analysis of **1** thus obtained with a chiral column revealed the optical purity to be >99% (Fig. 2).

As mentioned above, we have succeeded to develop the first method for the enantioselective synthesis of L^{-10} Bpa (1) from Z-L-Ser–OMe (5) used as the chiral synthon in 51.0% overall yield. This method is shorter in the reaction step and higher in the overall yield compared to the conventional methods for the synthesis of L-Bpa.¹⁵

The syntheses of the chiral fluorinated ¹⁰Bpa derivatives such as β -[4-(¹⁰B)borono-2-trifluoromethylphenyl]alanine and β -[4-(¹⁰B)borono-2,6-difluorophenyl]alanine¹⁶⁻¹⁸ using the present

Table 1

Yields of the Negishi coupling on various reaction conditions



method are currently being undertaken, and the results will be reported soon elsewhere.

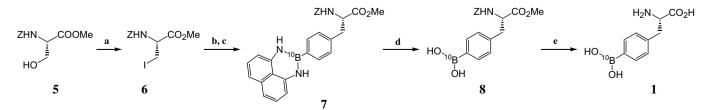
3. Experimental

3.1. General

All of the melting points are uncorrected and were measured by Yanaco MP-J3 (Yanaco Co., Ltd, Kyoto, Japan). Silica-gel column chromatography was carried out with silica gel PSQ100B (Fuji Silysia Chemical Ltd, Aichi, Japan). ¹H NMR spectra were measured on a Varian Mercury 300 (300 MHz, Varian Co., Ltd, USA) spectrometer. The chemical shifts in ¹H NMR are given in δ values from TMS used as the internal standard. HPLC analysis was performed on a LaChrom Elite (Hitachi high-Technologies Corporation, Tokyo, Japan) instrument equipped with a CROWNPAK AK CR(+) (5 μ m, 0.5×15 cm) (Daicel Chemical Industries, Ltd, Osaka, Japan) with a flow rate of 0.8 mL/min and detection at 230 nm. Elemental analyses were performed at the Micro Corder JM10 (J-Science Lab. Co. Ltd. Kvoto, Japan). Optical rotations were measured on a Jasco DIP-140 polarimeter (JASCO Co., Tokyo, Japan). ¹⁰B(OMe)₃ was purchased from Stella Chemifa Co. (Osaka, Japan), 1.2-Dibromoethane and *n*BuLi were purchased from Nacalai Tesque Inc. (Kyoto, Japan). 1,8-Diaminonaphtalene, TMS-Cl, and propylene oxide were purchased from Wako Pure Chemical Industries, Ltd (Osaka, Japan), 1,4-diiodo-benzene, tris(dibenzylideneacetone)dipalladium $[Pd_2(dba)_3]$ and tri(o-tolyl)phosphine $[P(o-tolyl)_3]$ were purchased from Sigma-Aldrich Co. (USA).

3.2. 4-[2,3-Dihydro-1*H*-naphtho[1,8-*de*]-1,3-diaza-2-(¹⁰B)-borinyl]iodobenzene (4)

To a solution of 1,4-diiodo-benzene ($\mathbf{2}$) (10.0 g, 30.3 mmol) in THF (100 mL) was added *n*-BuLi (1.6 M solution in hexane,



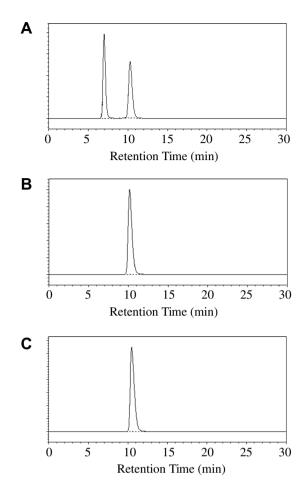


Figure 2. HPLC analysis of ¹⁰Bpa with a chiral column; (A) authentic sample of DL^{-10} Bpa, (B) authentic sample of L^{-10} Bpa, (C) synthetic L^{-10} Bpa. Analytical conditions-column: CROWNPAK AK CR(+); eluate: 0.4% aqueous HClO₄; flow rate: 0.8 mL/min; detection: UV 230 nm.

18.9 mL, 30.3 mmol) dropwise under cooling at -78 °C, and the solution was stirred for 30 min at -78 °C. To the reaction mixture was added ¹⁰B(OMe)₃ (3.75 g, 36.4 mmol) dropwise under cooling at -78 °C, and the solution was stirred for 5 h at -78 °C. To the reaction mixture was added saturated aqueous NH₄Cl (100 mL), and the solution was extracted with Et_2O (100 mL \times 3). The combined extracts were washed with H_2O (100 mL \times 3) and brine $(100 \text{ mL} \times 3)$, and dried over anhydrous MgSO₄. The dried extract was concentrated in vacuo, and the residue was dissolved in THF. To the solution was added 1,8-diaminonaphtalene (4.79 g, 30.3 mmol), and the mixture was stirred for 12 h. The reaction mixture was concentrated in vacuo, and the residue was purified by silicagel column chromatography (silica gel: 200 g, hexane/AcOEt = 9:1) to give **4** as brown crystals (7.04 g, 57.7%): mp 171–172 °C. ¹H NMR $(CDCl_3) \delta 5.98$ (s, 2H), 6.41 (dd, J = 7.5 Hz, 1.2 Hz, 2H), 7.05–7.17 (m, 4H), 7.37 (dd, J = 6.6 Hz, 1.8 Hz, 2H), 7.87 (dd, J = 6.6 Hz, 1.8 Hz, 2H). Anal. Calcd for $C_{16}H_{12}^{10}BIN_2$: C, 52.05; H, 3.28; N, 7.59. Found: C, 52.00; H, 3.60; N, 7.46.

3.3. N^{α} -Benzyloxycarbonyl-3-iodo-L-alanine methyl ester (6)

To a solution of I_2 (62.9 g, 248 mmol) and imidazole (16.9 g, 248 mmol) in CH₂Cl₂ (300 mL) was added PPh₃ (65.0 g, 248 mmol) portionwise, and the solution was stirred for 30 min at 0 °C. After stirring for 1 h at room temperature, to the reaction mixture was added the solution of **5** (31.5 g, 124 mmol) in CH₂Cl₂ (300 mL) dropwise, and the mixture was stirred for 5 h. The reaction mixture

was filtrated through the Celite, and the filtrate was washed with H₂O (300 mL × 3) and brine (300 mL × 3). The organic layer was dried over anhydrous MgSO₄, and concentrated in vacuo. The residue was purified by silica-gel column chromatography (silica gel: 400 g, hexane and then hexane/AcOEt = 9:1) to give **6** as colorless amorphous solid (37.2 g, 82.7%): mp 69–71 °C. $[\alpha]_D^{22} - 6.1(c 1.0, CHCl_3)$. ¹H NMR (CD₃Cl) δ 3.50 (d, *J* = 3.6 Hz, 2 h), 3.81 (s, 3H), 4.57–4.62 (m, 1H), 5.13 (s, 2H), 5.66 (d, *J* = 7.5 Hz, 1H), 7.28 (s, 5H). Anal. Calcd for C₁₂H₁₄INO₄: C, 39.69; H, 3.89; N, 3.86. Found: C, 39.92; H, 4.20; N, 3.86.

3.4. N^{α} -Benzyloxycarbonyl-*p*-[2,3-dihydro-1*H*-naphtho[1,8-*de*]-1,3-diaza-2-(¹⁰B)borinyl]-L-phenylalanine methyl ester (7)

A solution of 1.2-dibromoethane (310 uL, 3.60 mmol) in DMF (5 mL) was added to zinc dust (4.71 g. 72.0 mmol) under Ar atmosphere, and the suspension was stirred for 30 min at 90 °C. To the suspension was added TMS-Cl (129 µL, 3.60 mmol) at room temperature, and the mixture was stirred for 30 min. A solution of 6 (4.38 g, 12.0 mmol) in DMF (10 mL) was added to the reaction mixture, and the mixture was stirred for 1.5 h at 35 °C. To the reaction mixture were added 4 (4.01 g, 10.0 mmol), Pd₂(dba)₃ (274 mg, 0.30 mmol); and P(o-tolyl)₃ (304 mg, 1.00 mmol), and the mixture was stirred for 12 h at room temperature. The reaction mixture was filtered to remove the excess zinc dust, and the filtrate was diluted with AcOEt (100 mL). The solution was washed with H_2O (40 mL \times 3) and brine (40 mL \times 3), and dried over MgSO₄. The dried extract was concentrated in vacuo, and the residue was purified by silica-gel column chromatography (150 g, benzene and then benzene: acetone = 9:1) to give 7 as colorless crystals (4.00 g, 78.4%): mp 115–117 °C. $[\alpha]_D^{22} = 0.3$ (c1, MeOH). ¹H NMR (300 MHz, CDCl₃) δ 3.16 (ddd, J = 26.1 Hz, 14.4 Hz, 6.0 Hz, 2H), 3.75 (s, 3H), 4.62 (dd, J = 14.4 Hz, 6.0 Hz, 1H), 5.11 (s, 2H), 5.28 (d, J = 8.1 Hz, 1H), 6.00 (s, 2H), 6.41 (dd, J = 7.2 Hz, 1.2 Hz, 2H), 7.04–7.17 (m, 6H), 7.32–7.37 (m, 5H), 7.52 (d, J = 7.8 Hz, 2H). Anal. Calcd for $C_{28}H_{26}^{10}BN_3O_4 \cdot 0.5CH_3$ -COCH3: C. 69.81: H. 5.76: N. 8.28. Found: C. 69.54: H. 5.99: N. 8.47.

3.5. N^{α} -Benzyloxycarbonyl-*p*-(¹⁰B)borono-L-phenylalanine methyl ester (8)

To a solution of **7** (3.18 g, 6.23 mmol) in THF (75 mL) was added 2M H₂SO₄ (75 mL), and stirred for 2 h at room temperature. The reaction mixture was filtered, and the filtrate was extracted with AcOEt (50 mL × 3). The combined extracts were washed with 2M H₂SO₄ (50 mL × 3), saturated aqueous NaHCO₃ (50 mL × 3) and brine (50 mL × 3). The organic layer was dried over anhydrous MgSO₄, and concentrated in vacuo. The residue was purified by silica-gel column chromatography (100 g, CH₂Cl₂:MeOH = 19:1) to give **8** as colorless crystals. (2.10 g, 94.6%): mp 190–192 °C. $[\alpha]_D^{22} - 12.1(c1.00, MeOH)$. ¹H NMR (300 MHz, CD₃COCD₃) δ 3.00 (dd, *J* = 13.5 Hz, 9.0 Hz, 1H), 3.17 (dd, *J* = 13.5 Hz, 5.4 Hz, 1H), 3.67 (s, 3H), 4.49 (dd, *J* = 9.0 Hz, 5.4 Hz, 1H), 5.01 (s, 2H), 6.65 (d, *J* = 9.0 Hz, 1H), 7.14 (s, 2H), 7.24–7.37 (m, 7H), 7.04–7.17 (m, 4H), 7.80 (d, *J* = 7.8 Hz, 2H). Anal. Calcd for C₁₈H₂₀¹⁰BNO₆: C, 60.67; H, 5.66; N, 3.93. Found: C, 60.26; H, 6.03; N, 3.97.

3.6. p-(¹⁰B)Borono-L-phenylalanine (1)

A suspension of **8** (1.90 g, 5.33 mmol) in 3 M HCl (200 mL) was stirred for 17 h at 80 °C. The reaction mixture was washed with Et₂O (70 mL \times 3), and concentrated in vacuo. The residue was dissolved in *i*PrOH (50 mL), and to the solution was added propylene oxide (621 mg, 10.7 mmol). After stirring for 14 h, the reaction mixture was concentrated in vacuo. The crystalline residue was recrystallized from water to give **1** as colorless crystals (922 mg, 83.1%): mp 285–298 °C (decomp.). $[\alpha]_D^{25} - 5.2(c0.50, 1M \text{ HCl})$. ¹H NMR (300 MHz, D₂O) δ 2.97 (ddd, *J* = 18.2 Hz, 14.4 Hz, 5.4 Hz, 2H), 3.80 (dd, *J* = 14.4 Hz, 5.4 Hz, 1H), 7.15 (d, *J* = 7.8 Hz, 2H), 7.55 (d, *J* = 7.8 Hz, 2H). Anal. Calcd for C₉H₁₂¹⁰BNO₄·0.5H₂O: C, 49.76; H, 6.03; N, 6.45. Found: C, 49.90; H, 6.27; N, 6.23.

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